

Expression of Surface Molecules in Hematological Malignancies

Katarzyna Kotwica-Mojzych¹, Mariusz Mojzych²

Department of Histology, Embryology and Cytophysiology, Medical University of Lublin, Poland

Department of Chemistry, University of Siedlce, Poland

*E-mail: katarzynakotwicamojzych@umlub.pl

ABSTRACT

Hematological malignancies are represented in many cases by very aggressive tumors with poor outcome. For many years, immunophenotyping, that allows to describe specific surface antigens (also termed lineage-defining cell markers) in particular population of malignant cells has been widely used to assist in the diagnosis and monitoring of the broad spectrum of them. One of the greatest interest of both scientists and clinicians has been still multiple myeloma. Multiple myeloma is one the hematological neoplasm that belong to plasma cell dyscrasias. Their diagnosis is strictly associated with the description of specific phenotype on the population of malignant plasma cells. Among cell surface specific markers of these cells CD200, CD27, CD70 and CD40 are particularly significant. It was found that they directly associated with both immune escape and aggressiveness of malignant plasma cells. In these circumstances they may serve as a targets for a further treatment of multiple myeloma.

Keywords: hematological malignancies, phenotype, cell surface molecules, multiple myeloma, monoclonal antibodies

INTRODUCTION

Hematological malignancies are myeloid and lymphatic neoplasms caused by disruption of normal hematopoiesis in different stages. Many of them, despite being forefront of clinical interest, are characterized by poor outcome. Understanding their pathogenesis is still insufficient but discovery of lineage-defining cell surface markers significantly changed the paradigm of diagnosis and treatment. Among numerous hematological neoplasms one of the most curious from both scientific and clinical point of view is multiple myeloma.

Multiple myeloma is a hematological malignancy that is characterized by the clonal proliferation of malignant plasma cells in bone marrow, bone lesions, renal failure and immunodeficiency¹. Myelomagenesis is a complicated, comprehensive and multistep process that involves MGUS (Monoclonal Gammopathy of Undetermined Significance), SMM (Smoldering multiple myeloma) and symptomatic plasma cell myeloma also known as multiple myeloma². Among many targets in multiple myeloma the point of interest are the cell surface markers that define malignant plasma cell^{3,4,5}. Many years of work on the assessment of the phenotype of multiple myeloma cells allowed not only to specify but also to fully describe their characteristic surface markers. Among numerous of them, glycoproteins CD200, CD27, CD70 and CD40 deserve a special attention. Briefly, CD200 initially referred to as OX-2, is a highly conserved, 48 kDa type 1a transmembrane glycoprotein related to the B7 family of costimulatory receptors^{6,7}. It is widely expressed by most cells with the exception of skeletal muscle cells^{8,9,10,11}. Overexpression of CD200 in malignant plasma cells is associated with poorer outcome. Despite high-dose chemotherapy and ASCT, MM patients with cells overexpressing CD200 have shorter event freesurvival (EFS) when compared with patients whose cells do not (over) express CD200¹². In separate studies it has been reported that CD200 positive tumor cells can attenuate the ability of PBMCs to eradicate tumor cells¹³.

CD27, also termed as S152, is a 120-kDa type I transmembrane glycoprotein belonging to the nerve factor/TNF receptor gene family¹⁴. CD27 is expressed in normal plasma cells, but not malignant plasma cells, peripheral T cells, and a subset of mature B cells¹⁵. On binding to its ligand CD70, CD27 induces T dependent B cell immunity^{16,17}. As mentioned above, CD27 is expressed by normal plasma cells (CD19+CD38+) but its expression is reduced or absent in malignant plasma cells, a process which might have causal implications in progression of disease in MM^{18,19}. Thus low expression of CD27 is associated with clinically aggressive cases of multiple myeloma^{20,21}, and the lack of CD27 expression in plasmacytomas may be a marker for progression to myeloma (22). CD70, also referred to as CD27L, is a 50 kDa, homodimeric, type II-TNF related transmembrane glycoprotein^{23,24,25} which mediates the interaction between B- and T-lymphocytes and enhances the proliferation of activated T lymphocytes. Contrary to CD27, CD70 is overexpressed in malignant plasma cells and this might be associated with the increased proliferation of malignant plasma cells and their shorter overall survival. Interactions between CD27 and CD70 are known to promote the differentiation of peripheral blood memory B cells into plasma cells in association with several cytokines (IL-4, IL-10), CD40:CD40L signalling, and the up-regulation of PRDI-BF1/Blimp-1²⁶. Since CD70 is expressed on activated B and T cells it is possible that loss of CD27 during B cell differentiation in germinal centres (GC) contributes to immune escape and development of malignant clone of plasma cells^{18,27}. Restoration of the expression CD27 on malignant plasma cells would results its re-interaction with CD70, what could give a chance to induce apoptosis of these cells^{28,29}.

CD40L is a 32-33 kDa II type transmembrane protein belonging to the TNF-superfamily. It is expressed on the cell surface as one of three splice variants. Two shorter versions of the protein, 18 and 31 kDa respectively retain the ability to form trimers and bind to CD40 and thus can still apparently mediate biological signaling^{30,31,32}.

Overexpression of CD40L- CD40 axis in malignant plasma cells induce the cascade of intracellular signaling pathways responsible for its proliferation and survival^{33,34,35}. Preliminary studies showed that that targeting of the molecules mentioned above may be associated with significant beneficial effects. Preliminary studies showed that targeting CD200, CD70 and CD40L-CD40 axis may significantly enhance local immune response in tumor microenvironment. Namely, use of human anti-CD200 monoclonal in AML (acute myeloid leukemia) animal models successfully induced immune responses. They recovered the activity of NK (natural killer) cells³⁶.

Next, blockade of CD70 in variety CD70 positive cancer cell lines (e.g. multiple myeloma, Burkitt lymphoma, mantle cell lymphoma), in animal models, blocked their proliferation and survival and induce ADCC (antibody-dependent cell-mediated cytotoxicity), CDC (complement dependent cytotoxicity), and ADCP (antibody-dependent cell-mediated phagocytosis) mechanisms³⁷. Use of humanized anti-CD40 monoclonal antibody (SGN-40) against human MM.1S and MM1R multiple myeloma cell lines significantly inhibited their proliferation and survival, induced apoptosis and enhanced ADCC. SGN-40 both induced activity of FasL, TNF- α , and TRAIL and inhibited both AKT/I κ B α and ERK pathways. It is well known that IL-6R: IL-6 interactions promote bone loss, disease progression and drug resistance in multiple myeloma. SGN-40 diminished the expression of IL-6R³⁸.

OBJECTIVES

The first objective of this study was to present the significance of CD200, CD27, CD70 and CD40 in the development of multiple myeloma. The next purpose was to evaluate whether proper expression of CD200, CD27, CD70 and CD40 can put malignant plasma cells on the path to programmed type II death. The other aim of this study was to show if the above-mentioned molecules may become or direct or indirect therapeutic targets in the further treatment of multiple myeloma.

CONCLUSION/RESULTS

1. Reinstatement of normal CD27-CD70 axis may induce apoptosis of malignant plasma cells
2. Inhibition of CD200 and CD70 in malignant plasma cells may restore local immune response
3. inhibition of CD40 in positive malignant plasma cells both inhibits their proliferation and survival and induce apoptosis
4. CD200, CD27, CD70 and CD40 may serve as a future therapeutic targets.

REFERENCES

1. Kyle RA, Rajkumar SV. Multiple myeloma. *N Engl J Med* 2004; 351: 1860- 1873.
2. Korde N, Maric I. Myelomagenesis: Capturing Early Microenvironment Changes. *Semin Hematol.* 2011 ; 48 : 13-21
3. Leow CCY, Sze Yuan Low M. Targeted therapies for multiple myeloma. *J Pers Med.* 2021; 11:334
4. Zhou S. Targeted therapy of multiple myeloma. *Explor Target Antitumor Ther.* 2021; 2: 465-480
5. Ferguson ID et al. The surfaceome of multiple myeloma cells suggests potential immunotherapeutic strategies and protein markers of drug resistance. *Nat Commun.* 2022 ; 13 : 4121
6. Barclay AN. Different reticular elements in rat lymphoid tissue identified by localization of Ia, Thy-1 and MRC OX-2 antigens. *Immunology.* 1981 44: 727-736.
7. Barclay AN, Clark MJ, McCaughan GW. Neuronal/lymphoid membrane glycoprotein MRC OX-2 is a member of the immunoglobulin superfamily with a light-chain-like structure. *Biochem Soc Symp.* 1986; 51: 149-157.
8. Wright GJ, Cherwinski H, Foster-Cuevas M, Brooke G, Puklavec MJ, et al. Characterization of the CD200 receptor family in mice and humans and their interactions with CD200. *J Immunol.* 2003; 171: 3034-3046.
9. Kretz-Rommel A, Qin F, Dakappagari N, Cofield R, Faas SJ. Blockade of CD200 in the presence or absence of antibody effector function: Implications for anti-CD200 therapy. *J Immunol.* 2008; 180: 699-705.
10. Hoek RM, Ruuls SR, Murphy CA, Wright GJ, Goddard R, et al. Downregulation of the macrophage lineage through interaction with OX2 (CD200). *Science.* 2000; 290: 1768-1771.
11. Chen Z, Ma X, Zhang J, Hu J, Gorczynski RM. Alternative splicing of CD200 is regulated by an exonic splicing enhancer and SF2/ASF. *Nucleic Acid Research.* 2010; 38: 16684-96
12. Moreaux J, Hose D, Reme T, Jourdan E, Hundemer M, et al. CD200 is a new prognostic factor in multiple myeloma. *Blood.* 2006; 108: 4194-4197
13. Kretz-Rommel A, Qin F, Dakappagari N, Cofield R, Faas SJ. Blockade of CD200 in the presence or absence of antibody effector function: Implications for anti-CD200 therapy. *J Immunol.* 2008; 180: 699-705.
14. Bigler RD, Bushkin Y, Chiorazzi N. S152 (CD27). A modulating disulfidelinked T cell activation antigen. *J Immunol.* 1988; 141: 21-28.
15. Jung J, Choe J, Li L, Choi YS. Regulation of CD27 expression in the course of germinal center B cell differentiation: the pivotal role of IL-10. *Eur J Immunol.* 2000; 30: 2437-2443
16. Jacquot S. CD27/CD70 Interactions Regulate T Dependent B Cell Differentiation. *Immunol Res.* 2000; 21: 23-30.
17. Kobata T, Jacquot S, Kozlowski S, Agematsu K, Schlossman SF, et al. CD27-CD27 interactions regulate B-cell activation by T cells. *Proc Natl Acad Sci U S A.* 1995; 92: 11249-11253.
18. Katayama Y, Sakai A, Oue N, Asaoku H, Otsuki T, et al. A possible role for the loss of CD27-CD70 interaction in myelomagenesis. *Br J Haematol.* 2003; 120: 223-234.
19. Sakai A, Katayama Y, Otsuki T, Masuda R, Asaoku H, et al. Classification of myeloma cells by CD27. *Blood.* 2001; 98: 303b.
20. Zhan F, Hardin J, Kordsmeier B, Bumm K, Zheng M et al. Global gene expression profiling of multiple myeloma, monoclonal gammopathy of undetermined significance, and normal bone marrow plasma cells. *Blood.* 2002; 99: 1745-1757.
21. Guikema JE, Hovenga S, Vellenga E, Jelle JC, Wayel HA, et al. CD27 is heterogeneously expressed in multiple myeloma: low CD27 expression in patients with high-risk disease. *Br J Haematol.* 2003; 121: 36-43.
22. Morgan TK, Zhao S, Chang K, Haddix TL, Domanay E, et al. Low CD27 Expression in Plasma Cell Dyscrasias Correlates With High-Risk Disease. *Am J Clin Pathol.* 2006; 126: 545-551.
23. Goodwin RG, Alderson MR, Smith CA, Armitage RJ, VandenBos T, et al. Molecular and biological characterization of a ligand for CD27 defines a new family of cytokines with homology to tumor necrosis factor. *Cell.* 1993; 73: 447-456.
24. Tesselaar K, Gravestien LA, van Schijndel GMW, Borst J, van Lier RAW. Characterization of murine CD70, the ligand of the TNF receptor family member CD27. *J Immunol.* 1997; 159: 4959-4965.

25. Oshima H, Nakano H, Nohara C, Kobata T, Nakajima A, et al. Characterization of murine CD70 by molecular cloning and mAb. *Int Immunol.* 1998; 10: 517-526
26. Hoek RM, Ruuls SR, Murphy CA, Wright GJ, Goddard R, et al. (2000) Downregulation of the macrophage lineage through interaction with OX2 (CD200). *Science* 290: 1768-1771.
27. Bataille R, Jego G, Robillard N, Barille-Nion S, Amiot M, et al. (2006) The phenotype of normal, reactive and malignant plasma cells. Identification of “many and multiple myelomas” and of new targets for myeloma therapy. *Haematologica* 91: 1234-1240.
28. Prasad KV, Ao Z, Yoon Y, Wu MX, Rizk M, et al. CD27, a member of the tumor necrosis factor receptor family, induces apoptosis and binds to Siva, a proapoptotic protein. *Proc Natl Acad Sci USA.* 1997; 94: 6346-6351.
29. Xue L, Chu F, Cheng Y, Sun X, Borthakur A, et al. Siva-1 binds to and inhibits BCL-xL-mediated protection against UV radiation-induced apoptosis. *Proc Natl Acad Sci USA.* 2002; 99: 6925-6930.
30. Graf D, Muller S, Korthauer U, van Kooten C, Weise C, et al. A soluble form of TRAP (CD40 ligand) is rapidly released after T cell activation. *Eur J Immunol.* 1995; 25: 1749-1754.
31. Mazzei GJ, Edgerton MD, Losberger C, Lecoanet-Henchoz S, Graber P, et al. Recombinant soluble trimeric CD40 ligand is biologically active. *J Biol Chem.* 1995; 270: 7025-7028.
32. Ludwig B, Henn V, Schroder JM, Graf D, Krozcek RA. Induction regulation, and function of soluble TRAP (CD40 ligand) during interaction of primary CD41CD45RA1 T cells with dendritic cells. *Eur J Immunol.* 1996; 26: 3137-
33. van Kooten, Banchereau J. CD40-CD40 ligand. *J Leukoc Biol.* 2000; 67: 2-17.
34. Bradley JR, Poher JS. Tumor necrosis factor receptor-associated factors (TRAFs). *Oncogene.* 2001; 20: 6482 - 6491.
35. Shishodia S, Aggarwal BB. Nuclear factor-kappaB activation: a question of life or death. *J Biochem Mol Biol.* 2002; 35: 28-40.
36. Rastogi N, Baker S, Man S, Uger RA et al. Use of an anti-Cd200-blocking antibody improves immune responses to aml in vitro and in vivo. *Br J Haematol.* 2021; 193: 155-159
37. Silence K, Dreier T, Moshir M, Ulrichs P et al. ARGX-110, a highly potent antibody targeting CD70, eliminates tumors via both enhanced ADCC and immune checkpoint blockade. *MAbs.* 2014; 6: 523-532
38. Tai YT, Catley LP, Mitsiades CS, Burger R et al. Mechanisms by which SGN-40, a Humanized Anti-CD40 Antibody, Induces Cytotoxicity in Human Multiple Myeloma Cells: Clinical Implications. *Cancer Res.* 2004; 64: 2846-2852.